Introduction: Vulnerability of the CNS to metabolic and drug-related insults (Chapter 1)

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The CNS is particularly vulnerable to metabolic and drugrelated insults occurring along development. At the gestational period, the blood-brain barrier (BBB) provides protection against many insults affecting the mothers, or drugs taken by the mothers. Indeed, the BBB, as a barrier that excludes proteins, many charged or large molecules, and other substances present in the blood, is formed very early during development, probably, when the vessels first penetrate into the brain parenchyma and choroid epithelial cells differentiate.

Still, it has been suggested that the BBB of young animals is more permeable to small molecules than that in adults, either reflecting an immaturity of the tight junctions between the cerebral endothelial cells (Saunders and Møllgård, 1984; Møllgård and Saunders, 1986), or a greater metabolic demand by a rapidly growing tissue (Baños et al., 1978). Indeed, it has been reported that immature animals show a larger number of large pores in cerebral vessels and choroid plexus than that shown by more developed animals (Dziegielewska et al., 1979). In agreement, J. W. Olney first demonstrated that glutamate is toxic when administered subcutaneously to young animals (Olney, 1969), although it has been argued that the transport of excitotoxic amino acids to the brain by the BBB is low, and that the neurotoxicity of systemically administered glutamate is mainly found on cells of the arcuate nucleus, which is a circumventricular region outside the BBB (see Lefauconnier, 1992).

In the present chapter, Leon Navarro et al. (2005) present a paper investigating the effect of glutamate given by the drinking water to pregnant rats on metabotropic glutamate receptors assayed in the brain of mothers and pups, finding that the treatment produced a down-regulation of mGlu receptors and Gq/11 and phospholipase $C\beta1$

proteins in mothers, but not foetuses brains, supporting the idea that glutamate excitotoxicity does not primarily relate to BBB, but to increased endogenous viability. Glutamate is ubiquitously synthesized and released at high concentrations. In the extracellular space, glutamate is modulated by a powerful transport system to glia and nerve terminals, which, however, is very vulnerable to metabolic impairments, elevating extracellular glutamate to excitotoxic levels.

There are environmentally and synthetically produced amino acids that can penetrate the BBB over-stimulating glutamate receptors in the CNS. Kusama-Eguchi et al. (2005) show that 3-N-oxyalyl-L-2,3-diamino propanoic acid (L- β -ODAP; also known as β -N-oxalyl-amino-Lalanine [BOAA]), contained in Lathyrus Satirus grass pea, can accumulate in the spinal cord following parenteral administration, producing paraparesis of the hind legs in neonatal rats, associated to atrophy of the ventral root of the lumbar cord and degeneration of motor neurons. The mechanism by which L- β -ODAP is taken up and accumulated in the spinal cord has to be further investigated, because the incidence of the effect produced by L- β -ODAP is low. In humans, lathyrism was first described by the Greek physician Hippocrates and the Roman physician Galen, and was a major epidemic in Spain as recently as the 1940s (Spencer and Schaumburg, 1983). Lathyrism in humans is a slowly progressing irreversible spastic parapesis, disproportionately affecting young males. When >400 grams of L. sativus is consumed per day for several days (\sim 15–50 mg/kg/day), lathyrism can occur in just a few weeks (Cohn and Streifler, 1981). Pathologically, human lathyrism is thought to be attributable to selective loss of cortical Betz neurons specific to the leg region (Filiminoff, 1926), and in some ways the symptomatology resembles amyotrophic lateral sclerosis (Calne et al., 1986). BOAA has also been associated with degeneration of anterior horm cells in the spinal cord (Pearson and Nunn, 1981).

Propofol is a substituted phenol used for intravenous anaesthesia, producing, however, several side effects, including hallucination, sexual dis-inhibition and euphoria by, until now, unknown mechanisms. The hedonic and rewarding quality of these effects has suggested an action on mesolimbic dopamine circuitry, but seizures have also been reported, indicating an action on glutamatergic systems. These possibilities were investigated with in vivo microdialysis by Grasshoff et al. (2005), showing that propofol locally administered by a microdialysis probe decreases dopamine levels in ventral striatum, without affecting glutamate levels. Whether the decrease of dopamine levels relates to the side effects produced by propofol in humans remains to be investigated. Nevertheless, the methodological approach used by Grasshoff et al. (2005) is a competent method for dissecting in detail the action of a particular drug on specific neurocircuitry systems.

Perinatal asphyxia is still a serious clinical problem despite advances in perinatal and obstetric care, probably because of a lack of a suitable model for investigating the mechanisms by which short- and long-term consequences of perinatal asphyxia are produced. At the Karolinska Institutet, Stockholm, Sweden, a model for investigating perinatal asphyxia in the rat has been developed (Bjelke et al., 1991; Herrera-Marschitz et al., 1993), showing that dopamine (Chen et al., 1997a, b, c; Kohlhauser et al., 1999), as well as amino acid (Chen et al., 1997a; Kohlhauser et al., 1999; Engidawork et al., 2001) neurocircuitries are vulnerable to perinatal asphyxia. In the present chapter, Klawitter et al. (2005) show studies combining an in vivo and in vitro approach. After inducing perinatal asphyxia in vivo, surviving pups were used to prepare organotypic cultures, as shown by Morales et al. (2003), demonstrating that cultures prepared from asphyctic pups showed a decreased number of neurons labelled with an antibody against the NR1 subunit of the N-Methyl-D-aspartate (NMDA) receptor, and a decreased number of secondary to higher level branching of dopamine neurons labelled with an antibody against tyrosine hydroxylase.

The treatment of Parkinson's patients with the amino acid L-DOPA is well established, being an efficient treatment for rigidity and hypokinesia. L-DOPA is less effective against tremor, and does not reverse the course of the disease. Furthermore, the effectiveness of L-DOPA gradually declines by, until now, unknown mechanisms. It

has been suggested that L-DOPA's declining efficiency is due to impairment of L-DOPA transport and reduced biodisponibility in the targeted brain areas. The motor dyskinesia attending long-term L-DOPA has been attributed to the overexpression of striatal (i.e., basal ganglia) dopamine D₃ receptors, consequent to increased action of L-DOPA-induced production of brain-derived neurotrophic factor (BDNF) (Bezard et al., 2003; Bordet et al., 2000).

L-DOPA is transported into the brain by the L- transport system (Oldendorf, 1971), which is sodium independent and modulated by the availability of L-amino acids. The transport of L-DOPA competes with amino acids and proteins in the food. Furthermore, L-DOPA can be decarboxylated into dopamine and further de-aminated in the endothelial cells, before reaching the targeted areas. Thus, only a low percentage of systemically administered L-DOPA is transported into the brain. Indeed, as demonstrated by Zetterström et al. (1986), the amount of dopamine synthesized from a single clinically-relevant dose of L-DOPA reaches a tenth of the dopamine levels found in the striatal extracellular compartment of an intact brain, but that amount is enough to stimulate dopamine receptors rendered supersensitive following the lack of endogenous dopamine, and to revert the motor deficits.

In the targeted brain areas, L-DOPA can be taken up by the remaining dopamine nerve terminals, and further released as dopamine, or intracellularly metabolised by MAO dependent and/or independent mechanisms. It has been suggested that intracellular oxidation of L-DOPA or dopamine accelerates the progression of the disease (Fahn, 1997; Kostrzewa et al., 2000, 2002). In this chapter, however, Kostrzewa et al. (2005) discuss the idea that L-DOPA is actually neuroprotective, because the L-DOPA treatment reduces striatal tissue content of reactive oxygen species, and also induces BDNF release, which inturn enhances the expression of D3 receptors (Guillin et al., 2001), and perhaps neuritogenesis.

Thus, in this chapter several aspects related to the vulnerability of the CNS to metabolic and drug-related insults are discussed, enlightening the role of the BBB, CNS maturity for amino acid-induced excitotoxicity, and drug selectivity on particular neurocircuitries of the brain.

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References

- Baños G, Daniel PM, Pratt OE (1978) The effect of age upon the entry of some amino acids into the brain and their incorporation into cerebral protein. Dev Med Child Neurol 20: 335–346
- Bezard E, Ferry S, Mach U, Stark H, Leriche L, Boraud T, Gross C, Sokoloff P (2003) Attenuation of levodopa-induced dyskinesia by normalizing dopamine D3 receptor function. Nature Med 9: 762–767
- Bjelke B, Andersson K, Ogren SO, Bolme P (1991) Asphyctic lesion: proliferation of tyrosine hydroxylase-immunoreactive nerve cell bodies in the rat substantia nigra and functional changes in dopamine neurotransmission. Brain Res 543: 1–9
- Bordet R, Ridray S, Schwartz JC, Sokoloff P (2000) Involvement of the direct striatonigral pathway in levodopa-induced sensitization in 6hydroxydopamine-lesioned rats. Eur J Neurosci 12: 2117–2123
- Calne DB, Eisen A, McGeer E, Spencer PS (1986) Alzheimer's disease, Parkinson's disease, and motoneurone disease: abiotrophic interaction between ageing and environment. Lancet ii: 1067–1070
- Chen Y, Herrera-Marschitz M, Bjelke B, Blum M, Gross J, Andersson K (1997a) Perinatal asphyxia-induced changes in rat brain tyrosine hydroxylase-immunoreactive cell body number: effects of nicotine treatment. Neurosci Lett 221: 77–80
- Chen Y, Engidawork E, Loidl F, Dell'Anna E, Goiny M, Lubec G, Andersson K, Herrera-Marschitz M (1997b) Short- and long-term effects of perinatal asphyxia on monoamine, amino acid and glycolysis product levels measured in the basal ganglia of the rat. Dev Brain Res 104: 19–30
- Chen Y, Hillefors-Berglund M, Herrera-Marschitz M, Bjelke B, Gross J, Andersson K, von Euler G (1997c) Perinatal asphyxia induces long-term changes in dopamine D1, D2, and D3 receptor binding in the rat brain. Exp Neurology 146: 74–80
- Cohn DF, Streifler M (1981) Human neurolathyrism, a follow-up study of 200 patients, Part I. Arch Swisses Neurol Neurochem Psychiatry 128: 151–156
- Dziegielewska KM, Evans CAM, Malinowska DH, Møllgård K, Reynolds JM, Reynolds ML, Saunders NR (1979) Studies of the development of brain barrier systems to lipid insoluble molecules in fetal sheep. J Physiol (Lond) 292: 207–231
- Engidawork E, Loidl F, Chen Y, Kohlhauser S, Stoeckler S, Dell'Anna E, Lubec B, Lubec G, Goiny M, Gross J, Andersson K, Herrera-Marschitz M (2001) Comparison between hypothermia and glutamate antagonism treatments on the immediate outcome of perinatal asphyxia. Exp Brain Res 138: 375–383
- Fahn S (1997) Levodopa-induced neurotoxicity. Does it represent a problem for the treatment of Parkinson's disease? CNS Drugs 8: 376–393
- Filiminoff IN (1926) Pathologische-Anatomische Characteristik des Lathyrismus. Z Gesamte-Neurol Psychiatr 185: 76–92
- Guillin O, Diaz J, Carrol P, Griffon N, Scwartz J-C, Sokoloff P (2001) BDNF controls dopamine D3 receptor expression and triggers behavioural sensitization. Nature 411: 86–89
- Grasshoff C, Herrera-Marschitz M, Goiny M, Kretschmer BD (2005) Modulation of ventral pallidal dopamine and glutamate release by the intravenous anesthetic propofol studied by in vivo microdialysis. Amino Acids 28: 145–148
- Herrera-Marschitz M, Loidl CF, Andersson K, Ungerstedt U (1993) Prevention of mortality induced by perinatal asphyxia: hypothermia or glutamate antagonism? Amino Acids 5: 413–419

- Klawitter V, Morales P, Johansson S, Bustamante D, Goiny M, Gross J, Luthman J, Herrera-Marschitz M (2005) Effects of perinatal asphyxia on cell survival, neuronal phenotype and neurite growth evaluated with organotypic triple cultures. Amino Acids 28: 149–155
- Kohlhauser C, Kaehler S, Mosgoeller W, Singewald N, Kouvelas D, Prast H, Hoeger H, Lubec B (1999) Histological changes and neurotransmitter levels three months following perinatal asphyxia in the rat. Life Sci 64: 2109–2124
- Kostrzewa RM, Kostrzewa JP, Brus R (2000) Dopaminergic denervation enhances susceptibility to hydroxyl radicals in rat neostriatum. Amino Acids 19: 183–199
- Kostrzewa RM, Kostrzewa JP, Brus R (2002) Neuroprotective and neurotoxic roles of levodopa (L-DOPA) in neurodegenerative disorders relating to Parkinson's disease. Amino Acids 23: 57–63
- Kostrzewa RM, Nowak P, Kostrzewa JP, Kostrzewa RA, Brus R (2005) Peculiarities of L-DOPA treatment of Parkinson's disease. Amino Acids 28: 157–164
- Kusama-Eguchi K, Ikegami F, Kusama T, Suda A, Ogawa Y, Igarashi K, Watanabe K (2005) A rat model of neurolathyrism: repeated injection of L-β-ODAP induces the paraparesis of the hind legs. Amino Acids 28: 139–143
- Lefauconnier J-M (1992) Transport of amino acids. In: Bradbury MWB (ed) Physiology and pharmacology of the blood brain barrier. Springer, Wien New York, pp 117–150
- León Navarro D, Albasanz JL, Iglesias I, Ruiz MA, Martin M (2005) Effect of chronic glutamate administration to pregnant rats during gestation on metabotropic glutamate receptors from mothers and full-term fetuses brain. Amino Acids 28: 127–137
- Morales P, Klawitter V, Johansson S, Huaiquin P, Barros VG, Avalos AM, Fiedler J, Bustamante D, Gomez-Urquijo S, Goiny M, Herrera-Marschitz (2003) Perinatal asphyxia impairs connectivity and dopamine neurite branching in organotypic triple culture from rat substantia nigra, neostriatum and neocortex. Neurosci Lett 348: 175–179
- Møllgård K, Saunders NR (1986) The development of the human blood-brain and blood-CSF barriers. Neuropathol Appl Neurobiol 12: 337–358
- Oldendorf WH (1971) Brain uptake of radiolabeled amino acids, amines and hexoses after arterial injection. Am J Physiol 221: 1629–1639
- Olney JW (1969) Brain lesions, obesity and other disturbances in mice treated with sodium glutamate. Science 164: 719–721
- Pearson S, Nunn PB (1981) The neurolathyrogen, beta-N-oxalyl-L-alpha, beta-diaminopropionic acid, is a potent agonist at 'glutamate preferring' receptors in the frog spinal cord. Brain Res 206: 178–182
- Saunders NR, Møllgård K (1984) Development of the blood-brain barrier. J Dev Physiol 6: 45–57
- Spencer PS, Schaumburg HH (1983) Lathyrism: a neurotoxic disease. Neurobehav Toxicol Teratol 5: 625–629
- Zetterström T, Herrera-Marschitz M, Ungerstedt U (1986) Simultaneous measurement of dopamine release and rotational behaviour in 6-hydro-xydopamine denervated rats using intracerebral dialysis. Brain Res 376: 1–7

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